We want BETTER proteins for human applications.

Directed evolution is a molecular optimization process on a multi-dimensional fitness landscape, where fitness is performance and is defined by me.



Directed enzyme evolution: making use of smooth paths in the fitness landscape



Directed evolution can optimize an enzyme for a new job

















Enzymes (and other proteins) are highly evolvable

Proteins can adapt by accumulating beneficial mutations in a simple uphill walk.

New functions by changing tiny fraction (<2 %) of sequence.

Scary fact: many beneficial mutations are far from the active site and cannot be predicted, or even explained.



Directed evolution can optimize enzyme function in real time.



How can we access *novel* chemistry? (chemistry we have not found in the biological world)

Relevant to biology



Nature innovates all the time... atrazine





- 1950–1993 non-biodegradable; accumulated in soil
 - From 1993 onwards, fast degradation observed

Promiscuous activities can be a starting point for evolution of new functions





Second most abundant element in the Earth's crust

"...the literature is void of examples of biologically synthesized...silicon-carbon bonds."

- Silicon 2009, 1, 147



Inspiration from human chemistry? Brief history of transition metal catalysis for enantioselective carbene insertion into Si-H bonds



TTN = total turnover number

* = enantioselective catalyst

Evolution to explore/create the future...

Can an iron-heme protein do this?

Form a reactive Fe-carbenoid Intermediate



Transfer the carbene to a second substrate

Does it evolve?



Rhodothermus marinus cytochrome *c*

Gram-negative, thermohalophilic bacterium from hot springs in Iceland

124 amino acids, denaturation (melting) temperature $T_m = 106 \ ^{\circ}C$

Native function is electron transfer





1.23 Å X ray crystal structure of *Rma* cyt *c*

40-fold increased activity in just three generations



One enzyme: 20 example products, most are enantiopure





High resolution crystal structure shows a loop over the heme 'flipped' to form a new active site pocket





We can now MAKE carbon-silicon bonds with biology...can we BREAK them?

- Volatile methylsiloxanes (VMS) are the building blocks of silicone polymers
- \circ Not biodegradable, megatons/year production
- o Accumulating in the environment
- \circ Some are banned in the EU
- No enzyme known to break Si-C bonds





Linear siloxane examples:

 $\sum_{s_{i_0}, s_{i_1}}^{|} \sum_{s_{i_0}, s_{i_0}, s_{i_0}}^{|} \sum_{s_{i_0}, s_{i_0}, s_{i_0}}^{|}$

Cyclic siloxane examples:



Polydimethylsiloxane (PDMS):



Science 2024, 383, 438–443

RESEARCH

BIOCATALYSIS

Directed evolution of enzymatic silicon-carbon bond cleavage in siloxanes

Nicholas S. Sarai¹[†][‡], Tyler J. Fulton¹[†], Ryen L. O'Meara¹[†], Kadina E. Johnston²_§, Sabine Brinkmann-Chen¹, Ryan R. Maar³, Ron E. Tecklenburg³, John M. Roberts³, Jordan C. T. Reddel³, Dimitris E. Katsoulis⁴*, Frances H. Arnold¹*

Volatile methylsiloxanes (VMS) are man-made, nonbiodegradable chemicals produced at a megatonper-year scale, which leads to concern over their potential for environmental persistence, long-range transport, and bioaccumulation. We used directed evolution to engineer a variant of bacterial cytochrome P450_{BM3} to break silicon-carbon bonds in linear and cyclic VMS. To accomplish siliconcarbon bond cleavage, the enzyme catalyzes two tandem oxidations of a siloxane methyl group, which is followed by putative [1,2]-Brook rearrangement and hydrolysis. Discovery of this so-called siloxane oxidase opens possibilities for the eventual biodegradation of VMS. Since 2013, we have generated whole families of 'carbene transferases' with many activities not known in nature



Heme proteins can also generate and transfer nitrenes....









The enzyme can also selectively target the less reactive homobenzylic C–H bond



Todd Hyster, John McIntosh et al., J. Am. Chem. Soc. 136, 15505 (2014).

Cytochrome P411_{BM3} evolution for C-H insertion of nitrenes using <u>azide</u> precursors



see review Yang, Y.; Arnold, F. H. Acc. Chem. Res. 2021, 54, 1209-1225.

Intermolecular C(sp³)-H amination: unprotected primary amines using a <u>hydroxylamine ester</u> nitrene precursor





Zhijun Jia, Shilong Gao *JACS 142*, 10279 (2020)

The enzymes can aminate unactivated C-H bonds!





Soumitra Athavale, Shilong Gao, Anuvab Das JACS 2022

Evolution continues: P450s and hydroxylamine esters to access primary amines



Mao R. et. al. *Nat. Catal.* 2024 Athavale et. al. *JACS* 2022, *144*, 19097. Liu et. al. *JACS* 2022, *144*, 80. Amination enzymes use azides and hydroxylaminederived reagents.



Can enzymes use simple hydroxylamine?

Selective binding at N Higher N-O BDE



Hydroxylammonium chloride

- Bench stable
- Cheap
- Water is sole byproduct



Shilong Gao et al., JACS 145, 20196 (2023)





Das, A.; Gao, S. et al. J. Am. Chem. Soc. 2024, in press



Enzymatic activation of hydroxylamine





Analogous to peroxygenase? Activation of hydrogen peroxide by peroxygenase:



Unlike peroxygenase, NH₂OH aminating enzyme requires reducing environment

Gao, S.; Das, A., J. Am. Chem. Soc. 2023, 145, 20196.

This led to discovery of new aminating reagents: NO_2^- , NO, HNO all drive enzymatic amination



Das, A.; Gao, S. et al. J. Am. Chem. Soc. 2024, in press

Proposed mechanism inspired by nitrite reductase





Anuvab Das; Gao, S. et al. J. Am. Chem. Soc. 2024, in press



Derek Lowe (Opinion for *Science*):

"Here's one of those reactions that looks like alchemy to most of us in the synthetic organic community...

...these reactions really do look like magic, and they do chemistry that just can't be done with the reagents we have now. "



A heme protein is a self-assembling, DNA-encoded, chiral metal complex whose structure and electronic properties can be tuned by evolution.



Future: reducing the time and expense of directed evolution ("push the button")

Improving directed evolution with machine learning



All these steps can be automated....



A fully autonomous system for protein engineering/evolution (P. Romero, U. Wisconsin)



Rapp et al., Self-driving laboratories to autonomously navigate the protein fitness landscape. Nat Chem Eng 1, 97–107 (2024).